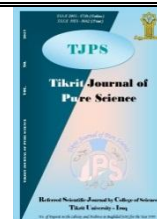




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Ecology, Morphology and Molecular Confirmation of *Chaetogaster limnaei* (Annelida: Naididae) retrieved from freshwater snail *Physa acuta* from Greater Zab River, Iraq.

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ABSTRACT

The present study is concerned with the *Chaetogaster limnaei* retrieved from the freshwater snail *Physa acuta* collected from (March-October 2021) from various locations near Chama-dubz village on the Greater Zab River, Erbil province, Iraq. Some Physicochemical variables, like water temperature, pH, Dissolved Oxygen, and Calcium ion experiment on a freshwater snail. The result showed statistically significant differences among Monthly variations with variables (number of snails, water temperature, pH and DO, and Calcium ion). In addition, isolated the *Chaetogaster limnaei* from *Physa acuta* with a prevalence was 5.75% (33/574), which has no dorsal and ventral chaeta approximately 14 to 20 per bundle, lack in segments 3 to 5, and chaetae and a vestige of a prostomium with sharply bent teeth. The identified morphologically and molecularly confirmed by using a COI sequence marker confirmed a 100% match to species with accession number (KF952336.1). As a result, the current study is the first report of *Chaetogaster limnaei* in Iraq, supported by phylogenetic analysis using Maximum Likelihood and Maximum Parsimony Methods.

البيئة، المظهر الخارجي والتأكيد الجزيئي لـ *Chaetogaster limnaei* (Annelida: Naididae)

المسترجعة من حلزون المياه العذبة *Physa acuta* من نهر الزاب الكبير ، العراق

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المخلص

تتناول الدراسة الحالية *Chaetogaster limnaei* المستخرج من حلزون المياه العذبة *Physa acuta* الذي تم جمعه من (مارس - أكتوبر 2021) من مواقع مختلفة بالقرب من قرية Chama-dubz على نهر الزاب الكبير ، محافظة أربيل، العراق. بعض المتغيرات الفيزيائية والكيميائية، مثل درجة حرارة الماء، ودرجة الحموضة، والأكسجين المذاب، وأيون الكالسيوم تجرب على حلزون المياه العذبة. أظهرت النتائج وجود فروق ذات دلالة إحصائية بين المتغيرات الشهرية مع المتغيرات (عدد القواقع ، درجة حرارة الماء ، الأس الهيدروجيني و DO ، وأيون الكالسيوم). بالإضافة إلى ذلك ، تم عزل الجيني COI. ونتيجة لذلك ، فإن الدراسة الحالية هي أول تقرير لـ *Chaetogaster limnaei* في العراق ، مدعوماً بتحليل النشوء والتطور باستخدام طرق الحد الأقصى من الاحتمالية Maximum Likelihood والحد الأقصى من Maximum Parsimony.

1. Introduction

Chaetogaster limnaei is an oligochaete considered ectosymbiont and cosmopolitan species belonging to the family Naididae, its close commensal relationship with more than 40 species of freshwater snails, mostly memberships of Physidae, Planorbidae and Lymnaeidae [1-3], infesting the mantle, pulmonary cavities, or foot mucus [4]. *C. limnaei* has been reported in association with a variety of host taxa across the world including Africa, Asia, Australia, Europe, North America, and South America [5].

External morphological characteristics including chaetae, proboscis, and gills serve to distinguish the genus *Chaetogaster*. The chaetae, which include the segment on which dorsal chaetae originate, the number of chaetae per bundle, the relative sizes of bifid chaetae teeth, as well as the existence or lack of hair chaetae, are regarded particularly essential features for the identification of genera and species of Naidinae [6, 7]. Furthermore, the difficult taxonomic based on morphological characteristics compromises, then using DNA barcodes like mitochondrial Cytochrome Oxidase I (COI) primer serves as an efficient barcode for oligochaetes, making it easier to use them in biomonitoring and advancing ecological diagnostics [8, 9].

In Iraq, various species of *Chaetogaster* were recorded, including *C. limnaei* by authors in different localities [7, 10-12]. Previously, not recorded in the Greater Zab river only one paper by Ali [13] reported another species *Chaetogaster langi*.

In this research, we studied the impacts of environmental variables on snails and their *C. limnaei*, then considered to be the first report of *C. limnaei* invasive *Physa acuta* in Greater Zab river, Iraq, which were supported by morphology and molecular (COI) primer, added to Iraqi Oligochaeta fauna.

2. Material and method

Study area

Species of living freshwater snail (*Physa acuta*) collected from various sites in Iraqi Greater Zab River, Erbil Province close to Chama-dubz Village, during (March-October/ 2021) (36°13'56.2"N 43°37'30.6"E), were transferred to the Advanced Invertebrate Lab, Education College, Salahaddin University, in a container filled with river water.

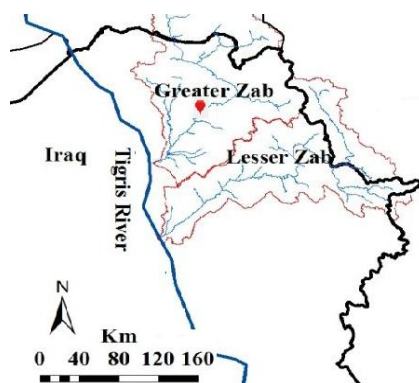


Fig. 1: Studied area on Greater Zab river (North Iraq) [14].

In the field, water temperature and pH conductivity were directly evaluated utilizing portable HANNA instruments, while dissolved oxygen and calcium ions content were measured in the laboratory, all variables performed depend on [15]. The species identified morphology by using the key for identification [16]. A traditional method was employed to isolate the naididae species from snails, which involved dissection of the snails (crushing infected snails) [3] and preserving in (90%) ethanol for morphology study and molecular identification.

DNA Extraction, COI Amplification, and Sequencing

The Oligochaete DNA fixed in "Ethanol %90", centrifuged at 8,000 RPM/3min., and the solution "ethanol" eliminated. A commercially available kit (GeneAll® Exgene™ Clinic SV, Korea) was used to extract the DNA following the manufacturer's protocol. The DNA barcoding for oligochaete was achieved based on amplification and sequencing of the mitochondrial DNA Cytochrome Oxidase subunit I (COI) primer (680-720 bp long) Folmer [17] by using forward primer LCO1490 (GGTCAACAAATCATAAAGATATTGG) and reverse primer HCO2198 (TAAACTTCAGGGTGACCAAAAAATCA). The PCR Coctail (40 µl) comprised of: 20 µl master mix, 4 µl "F & R primers", 3 µl DNA template, and 13 µl (ddH₂O). PCR program running under 94 C°/3 min; 35 cycles of 94 C°/60 sec; 50 C°/45 sec; 72 C°/45 sec, and 72 C°/7 min. in MJ Research, Applied Biosystem (AB) thermocycler. PCR results were separated in a 1.3% agarose gel and dyed with a UV-safe dye to detect DNA [14]. DNA sequencing was performed by utilizing a Macrogen Inc. ABI 3730XLs nucleotide sequencing analyzer (Korea). All the DNA sequences were edited and aligned with (ClustalW algorithm), available in MUSCLE program within EMBL-EBI (<https://www.ebi.ac.uk/Tools/msa/muscle/>). All sequences were subjected to the Basic Local Alignment Search Tool for nucleotides "Blastn" implemented in the NCBI GenBank database. Further estimates of COI variation and relationships among species use the Maximum Parsimony (MP) analysis, Maximum Likelihood (ML) method, and Kimura 2-parameter model [18]. The index of consistency is (0.867188), the index of retention is (0.892405), and the index of composite is (0.773883) for all locals and parsimony-informative sites. In the bootstrap test (100 replicates), the proportion of duplicate trees in which the linked taxa clustered together is displayed next to the branches [19]. The MP tree was obtained using the Subtree-Pruning-Regrafting (SPR) algorithm [20], and in the ML method, Branches corresponding to partitions reproduced in less than 50% bootstrap replicates are collapsed. All positions with less than 100% site coverage were eliminated. Evolutionary analyses were

conducted in MEGA X [21] including a sequence representative from species currently available on GenBank.

Statistical analysis

The data collected were subjected to using the one-way (ANOVA) to analyze the impact of months and Physico-chemical parameters, and Tukey's multiple comparison test to compare environmental between them.

3. Results and Discussion

3.1. Results

3.1.1. Ecology description

Several Physico-chemical variables have been researched on *P. acuta* (Table 1). Monthly variation statistically showed significant differences ($p < 0.05$) with all parameters (water temperature, DO, and Ca^{+2}) (Fig. 3). A total of (574) freshwater snails *P. acuta* were collected, 33 of them were infected with *C. limnaei*, with a prevalence (5.75 %). The maximum number of snails examined recorded in July and minimum in March (Fig. 4). While the highest snail infected during both months June and July and the lowest in March was zero (Fig. 4).

3.1.2. Morphology description

It has no dorsal and ventral chaeta, approximately 14 to 20 per bundle are lack in segments 3 to 5, and the existence of an inconspicuous prostomium. A chaetae and a vestige of a prostomium with sharply bent teeth are further characteristics of the species (Fig. 2).

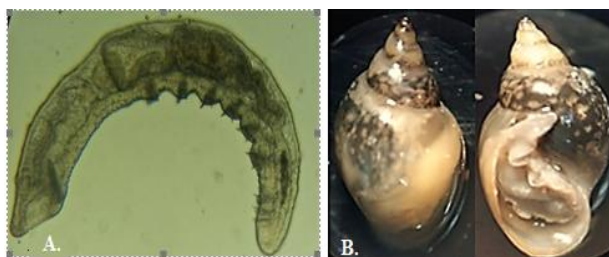


Fig. 2: A. *Chaetogaster limnei*, lateral view (200X); B. *Physa acuta*, dorsal and ventral view (20 X).

3.1.3. Molecular result

The GenBank Blasted DNA (COI) sequence of *C. limnaei* confirmed a 100% match to species with accession number (KF952336.1) (Fig. 6).

Table 1: Number of snails examined (574) versus parameters of Greater Zab River, represents as Mean \pm SE, and P-value from March to September 2021

Parameters	Mean \pm SE Min.-Max.	P-value	Significant
No. snail infected	4.12 \pm 1.2 0-9	<0.0001	****
Water Temp. (°C)	25.99 \pm 2.06 17-32	0.0219	*
pH	7.83 \pm 0.05 7.6-8.1	0.9709	NS
DO (mg/L)	8.42 \pm 0.71 6.1-11.1	0.0075	**
Calcium Con. (mg/L)	42.79 \pm 1.21 39.75-49	0.080	**

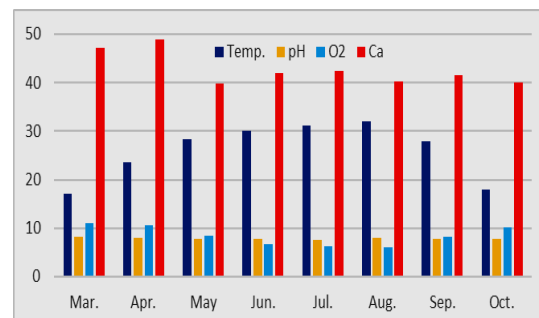


Fig. 3: Monthly variation vs. Water Temperature, pH, O₂, and Ca²⁺ of Greater Zab river in the studied site

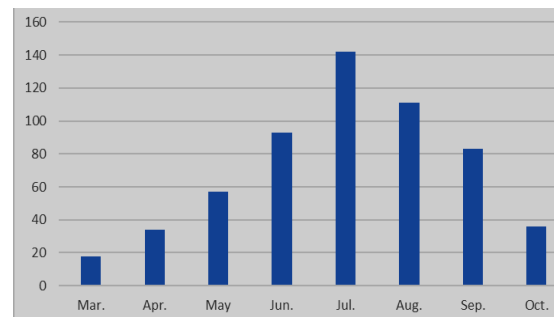


Fig. 4: Monthly variation vs. Number of snails examined

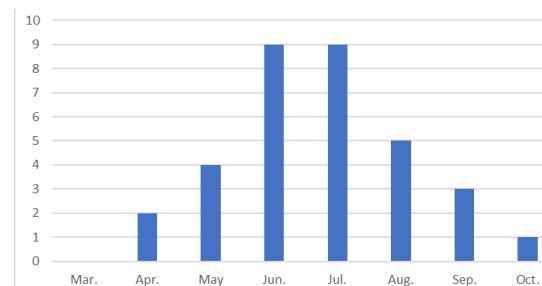


Fig. 5: Monthly variation vs. Number of snail infected

3.2. Discussion

The current investigation identified *Chaetogaster limnaei* isolated from *Physa acuta* in Greater Zab River (north Iraq). Monthly variations show significantly with snails *Physa acuta* (Table 1), the minimum and maximum numbers examined ranged (18-142) during March and July (Fig. 4), because this snail invades all freshwaters in the world and is present in a moderate amount of aquatic vegetation and organic debris [22, 23]. Another explanation is that it is hermaphrodite, which means it can self-fertilize and generate a high number of offspring throughout the year [24]. Also, the highest prevalence of infected snails with oligochaetes was (9.36 %) observed in July, and the lowest in March (0%). Statistically, shows significant differences ($p < 0.05$) between *Physa acuta* with water temperature, DO, and Ca^{+2} and non-significant with pH (Table 1). In Figure (4), shows increasing the number of snails gradually, this result agrees with Sahin [25] claimed that the high adaptation capabilities of gastropods show a wide distribution of areas, common in organically-rich polluted waters can tolerate low levels of oxygen values. Furthermore, it is found in warm water and huge numbers throughout the summer, which is likely due to the appropriate temperatures for breeding and

reproduction during these months., these results are in line with [26-28]. High temperatures in the snail vectors cause thermal stress [29]. The pH value ranges (7.3-8.1) during the most studied period (Fig. 3), and monthly variation agree with Iraqi inland water is considered neutral to be alkaline, reflecting the geologic region [30]. The Dissolved oxygen content of water, which is critical for all living creatures [31]. It ranged from 6.5-12.3 mg / L (Fig. 3), and this outcome was comparable to Ali [32], and greater than that recorded by Shekha [33] at the same location. The lower DO level was detected during warm months, due to an increase in water temperature (32 C°) due to raising in organic waste disposal, as well as an increase in decomposer bacteria activity [13]. The high DO level was recorded in March, this was probably linked to water temperature and the dilution effect of rainfall [34]. The calcium ion is one of the most significant components in the snail shell [35], the ranges (30.80-50.02 80 mg/l) were noted in the spring season, and this phenomenon came by the results [13, 36]. Dalesman and Lukowiak [37] claimed that snails exposed to lower level (20 mg/l) had significantly slower locomotion than snails exposed to a higher levels (80 mg/l). In addition, various environmental factors have a collective impact on organisms, therefore it is challenging to separate the effects of different factors [38]. Additionally, it has been demonstrated that aquatic vegetation affects snails' habitat as a result of elements like food availability, competition, and interactions between predators and prey [39].

The molecular technique proved effective in verifying the precision with which a species was identified depend on morphology. A fragment of 597 base pairs produced by COI gene amplification in the *C. limnaei* specimen. Analysis of BLASTn revealed that 100% identical with *C. limnaei* (KF952336.1) from New York, USA [1] (Fig. 6). In both trees our sequence of *C. limnaei* formed a monophyletic group (100% MP and 99% ML), together with *C. limnaei* from New York, USA, within a clade (66% MP and 63% MP) comprised of further *Chaetogaster* species, and other species *Chaetogaster diaphanous* (LN810268.1) by

Vivien et. al, [9] in Switzerland and (JQ519897.1) by Envall et. al, [40] in from Europe, North America, and China were located in another clade, for which relationship supported with *Chaetogaster cf. diastrophus* (MK837023.1) by Klinth et. al, [41] in Iceland, the out-group is *Lumbriculus variegatus* (FJ639308.1) by Gustafsson et. al, [42].

In the current investigation, detected that *C. limnaei* and *P. acuta* were commensal, and an assessment of the freshwater snails revealed any destruction to the paleal tissues, suggesting that this is a theory that needs additional research. In contrast, assumed that an epizotic antibiosis relationship between organisms at high infestation levels proposed by the decreased growth levels of the infected snails. *C. limnaei* is an omnivorous commensal that consumes a wide range of prey depend on particle size, includes: rotifers, foraminiferans, ciliates, flagellates, diatoms, filamentous algae, and trematode cercariae [5].

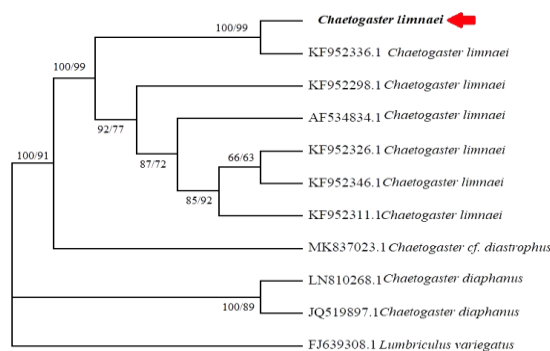


Fig. 6: The history of evolutionary by using the Maximum Parsimony analysis and Maximum Likelihood method and Kimura 2-parameter model “Numbers above nodes represent MP probabilities, and numbers below are ML bootstrap values”.

Conclusion

It can be concluded, based on the outcomes of this investigation, considered to be the first report of *C. limnaei* invasive *Physa acuta* in Greater Zab river, Iraq, which were supported by morphology and molecular (COI) primer, added to Iraqi Oligochaeta fauna.

References

- [1] Smythe, A.B., Forgrave, K., Patti, A., Hochberg, R. & Litvaitis, M.K. (2015). Untangling the ecology, taxonomy, and evolution of *Chaetogaster limnaei* (Oligochaeta: Naididae) species complex. *The Journal of Parasitology*. **101** (3): 320-326.
- [2] Timm, T. & Martin, P.J. (2015). Clitellata: Oligochaeta, In Thorp and Covich's Freshwater Invertebrates, Thorp, J.H. and Rogers, D.C., Academic press, Elsevier: 529-549.
- [3] Collado, G.A., Cabrera, F.J., Ballesteros, G.I., Villalobos, N.I. & Aguayo, K.P. (2019). First report of *Chaetogaster limnaei* (Annelida: Naididae) in Chile based on samples retrieved from an invasive freshwater snail. *Journal Revista Mexicana de Biodiversidad*. **90** (1): e90257.
- [4] Ibrahim, M.M. (2007). Population dynamics of *Chaetogaster limnaei* (Oligochaeta: Naididae) in the field populations of freshwater snails and its implications as a potential regulator of trematode larvae community. *Journal of Parasitology research*. **101** (1): 25-33.
- [5] Stoll, S., Früh, D., Westerwald, B., Hormel, N. & Haase, P. (2013). Density-dependent relationship between *Chaetogaster limnaei* (Oligochaeta) and the freshwater snail *Physa acuta* (Pulmonata). *Freshwater Science*. **32** (2): 642-649.
- [6] Timm, T. (2009). A guide to the freshwater Oligochaeta and Polychaeta of Northern and Central Europe, Lauterbornia: 235pp.

- [7] Jaweir, H.J. & Radhi, M.M. (2013). Naididae (Clitellata: Oligochaeta) and Aeolosomatidae (Polychaeta: Aphanoneura) species associated with aquatic plants in Tigris river/Baghdad/Iraq. *Baghdad Science Journal*. **10** (1): 116-125.
- [8] Kvist, S., Sarkar, I.N. & Erséus, C. (2010). Genetic variation and phylogeny of the cosmopolitan marine genus Tubificoides (Annelida: Clitellata: Naididae: Tubificinae). *Molecular Phylogenetics and Evolution*. **57** (2): 687-702.
- [9] Vivien, R., Wyler, S., Lafont, M. & Pawlowski, J. (2015). Molecular barcoding of aquatic oligochaetes: implications for biomonitoring. *PloS one*. **10** (4): e0125485.
- [10] Al-Kuti, S. (2000). Use of annelid, oligochaeta as bioindicators in evaluating the pollution in Al-Dewanla River. MSc. Thesis, College of Education, Univ. of Al-Qadisiyah, Iraq: 132 pp.
- [11] Al-Abbad, M. (2009). Identification and Biology of the species *Chaetogaster limnaei* von Baer 1827 (Oligochaeta: Naididae) isolated from some Basrah marshes snails in the south of Iraq. Ph. D. thesis, University of Basrah, Iraq: 145 pp.
- [12] Jaweir, H.J. (2014). Checklist of aquatic oligochaetes species in Tigris–Euphrates river basin. *Baghdad Science Journal*. **11** (3): 1397-1404.
- [13] Ali, L. (2007). A study of macroinvertebrates community in the middle sector of Greater Zab River, Iraq. Ph. D. Thesis. University of Baghdad, Iraq: 123 pp.
- [14] Bashe, S.K. & Ali, L.A. (2019). First molecular identification and phylogenetic tree of *Petasiger exaeretus* Dietz, 1909 (Digenea: Echinostomatidae) from an intermediate host *Radix auricularia* (L., 1758) in Greater Zab river, Iraq. *Annals of Agricultural and Environmental Medicine*. **26** (3): 504-507.
- [15] A.P.H.A. (2012). Standard Methods for the Examination of Water and Waste Water, Washington, USA: 1360pp.
- [16] Mansoorian, A. (2001). Freshwater Gastropod of Khuzestan Province, South-West Iran. *Iranian International Journal Science*. **2** (2): 96-103.
- [17] Folmer, O., Black, M., Hoeh, W., Lutz, R. & Vrijenhoek, R. (1994). DNA primers for amplification of mitochondrial Cytochrome C Oxidase subunit I from diverse metazoan invertebrates *Molecular and Marine Biology and Biotechnology*. **(3)**: 294–299.
- [18] Kimura, M. (1980). A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *Journal of molecular evolution*. **16** (2): 111-120.
- [19] Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution* **39** (1): 783-791.
- [20] Nei, M. & Kumar, S. (2000). Molecular evolution and phylogenetics, Oxford University Press: 122pp.
- [21] Kumar, S., Stecher, G., Li, M., Knyaz, C. & Tamura, K. (2018). MEGA X: molecular evolutionary genetics analysis across computing platforms. *Molecular biology and evolution*. **35** (6): 1547–1549.
- [22] Wethington, A.R. & Lydeard, C. (2007). A molecular phylogeny of Physidae (Gastropoda: Basommatophora) based on mitochondrial DNA sequences. *Journal of Molluscan Studies*. **73** (3): 241-257.
- [23] Al-Waaly, A.B. (2014). Use of molecular technique and scanning electron microscope in freshwater snails taxonomy and their infection with larval trematoda in the middle and south of Iraq. Al-Qadisiya University, Iraq: 171 pp.
- [24] Maqboul, A., Aoujdad, R., Fadli, M. & Fekhaoui, M. (2014). Population dynamics of *Physa acuta* (Mollusca: Pulmonata) in the lakes of Rif mountains (northern Morocco, Ouergha watershed). *Journal of Entomology and Zoology Studies*. **2**:240-245.
- [25] Sahin, S.K. (2012). Gastropod species distribution and its relation with some physico-chemical parameters of the Malatya's streams (East Anatolia, Turkey). *Acta Zoologica Bulgarica*. **64** (2): 129-134.
- [26] Ali, A., Aziz, N. & Hamza, H. (2007). Abundance, occurrence, seasonal changes and species composition of Macroinvertebrates in the restored Iraqi southern marshes. *Marsh Bulletin*. **2** (1): 80-95.
- [27] Al-Abbad, M., Salman, S.D. & Al-Qarooni, I. (2015). Biodiversity of the macroinvertebrates in the Southern Iraqi Marshes, with a special reference to oligochaeta. *Journal of Biodiversity and Environmental Sciences*. **7** (1): 61-71.
- [28] Mahmoud, A., Fangary, H., Hussein, M. & Obuid-Allah, A. (2011). Population dynamics of freshwater snails (Mollusca: Gastropoda) at Qena Governorate, Upper Egypt. *Egyptian Academic Journal of Biological Sciences, B. Zoology*. **3** (1): 11-22.
- [29] Njoku-Tony, R. (2007). Ecological studies on some human and animal trematodes in parts of Imo State, Nigeria. Ph.D Thesis, Imo State University, Owerri, Imo State, Nigeria: 130 pp.
- [30] Maulood, B., Hinton, G. & Al-Dosky, H. (1980). A study on the blue green algal flora of Arbil province, Iraq. *Zanco. Series A. Pure and Applied Sciences (Iraq)*. **6** (2): 67-89.
- [31] WHO (2006). Guidelines for the safe use of wastewater, excreta and greywater, World Health Organization. Available from: <https://apps.who.int/iris/handle/10665/78265>: 101pp.
- [32] Ali, L.A. (2010). Seasonal Variation in Physico-Chemical Properties and Zooplankton Biomass in Greater Zab River, Iraq. *Jordan Journal of Biological Sciences*. **147** (615): 1-12.
- [33] Shekha, Y.A. (2008). The effect of Erbil city wastewater discharge on water quality of Greater Zab river, and the risks of irrigation. Ph.D Thesis, University of Baghdad, Iraq.: 169 pp.
- [34] Telang, S. (1990). Effects of reservoir-dam, urban, industrial, and sewage treatment run-off on the presence of oxygen and organic compounds in the Bow River. *Water, Air, and Soil Pollution*. **50** (1-2): 77-90.
- [35] Soído, C., Vasconcellos, M.C., Diniz, A.G. & Pinheiro, J. (2009). An improvement of calcium determination technique in the shell of molluscs.

Brazilian Archives of Biology and Technology. **52** (1): 93-98.

[36] Saadalla, H. (1998). Ecological study on the effect of Himreen impoundment on the benthic and planktonic invertebrates of river Diyala. Ph. D. Thesis University of Baghdad, Iraq.: 254 pp.

[37] Dalesman, S. & Lukowiak, K. (2010). Effect of acute exposure to low environmental calcium on respiration and locomotion in *Lymnaea stagnalis* (L.). *Journal of Experimental Biology.* **213** (9): 1471-1476.

[38] Berrie, A. (1970). Snail problems in African schistosomiasis, In *Advances in Parasitology*, Dawes, B., Academic Press (Inc.), London, Elsevier: 43-96.

[39] Usman, A., Babeker, E. & Malann, Y. (2017). Effects of some physico-chemical parameters on prevalence of intermediate host of animal trematodes in

Bauchi State, Nigeria. *Science World Journal.* **12** (4): 94-97.

[40] Envall, I., Gustavsson, L.M. & Erseus, C. (2012). Genetic and chaetal variation in Nais worms (Annelida, Clitellata, Naididae). *Zoological Journal of the Linnean Society.* **165** (3): 495-520.

[41] Klinth, M., Kreiling, A.-K. & Erséus, C. (2019). Investigating the Clitellata (Annelida) of Icelandic springs with alternative barcodes. *Fauna norvegica.* **39**:119-132.

[42] Gustafsson, D.R., Price, D.A. & Erséus, C. (2009). Genetic variation in the popular lab worm *Lumbriculus variegatus* (Annelida: Clitellata: Lumbriculidae) reveals cryptic speciation. *Molecular Phylogenetics and Evolution.* **51** (2): 182-189.